

Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 17 (2007) 5595-5599

Dihydroxypyridopyrazine-1,6-dione HIV-1 integrase inhibitors

John S. Wai,^{a,*} Boyoung Kim,^a Thorsten E. Fisher,^a Linghang Zhuang,^a Mark W. Embrey,^a Peter D. Williams,^a Donnette D. Staas,^a Chris Culberson,^a Terry A. Lyle,^a Joseph P. Vacca,^a Daria J. Hazuda,^b Peter J. Felock,^b William A. Schleif,^c Lori J. Gabryelski,^c Lixia Jin,^d I-Wu Chen,^d Joan D. Ellis,^d Rama Mallai^e and Steven D. Young^a

^aDepartment of Medicinal Chemistry, Merck Research Laboratories, West Point, PA 19486, USA
 ^bDepartment of Biological Chemistry, Merck Research Laboratories, West Point, PA 19486, USA
 ^cDepartment of Vaccine and Biologics Research, Merck Research Laboratories, West Point, PA 19486, USA
 ^dDepartment of Drug Metabolism and Pharmacology, Merck Research Laboratories, West Point, PA 19486, USA
 ^eDepartment of Drug Metabolism, Merck Research Laboratories, Rahway, NJ 07065, USA

Received 29 June 2007; revised 26 July 2007; accepted 27 July 2007 Available online 22 August 2007

Abstract—A series of potent novel dihydroxypyridopyrazine-1,6-dione HIV-1 integrase inhibitors was identified. These compounds inhibited the strand transfer process of HIV-1 integrase and viral replication in cells. Compound 6 is active against replication of HIV with a CIC $_{95}$ of 0.31 μ M and exhibits no shift in potency in the presence of 50% normal human serum. It displays a good pharmacokinetic profile when dosed in rats and no covalent binding with microsomal proteins in both in vitro and in vivo models. © 2007 Elsevier Ltd. All rights reserved.

Human immunodeficiency virus-type 1 (HIV-1) is the etiological agent of acquired immunodeficiency syndrome (AIDS). The unique nature of the replicative cycle of HIV-1 provides many potential targets for chemotherapeutic intervention. One of these, the viral enzyme integrase, catalyzes the insertion of the proviral DNA into the genome of host cells. Integration is a multistep process which includes three different biochemical steps: assembly of the proviral DNA on integrase, endonucleolytic processing of the proviral DNA, and strand transfer of the proviral DNA to host cell DNA.

1,3-Diketoacids 1 were reported to be effective integrase inhibitors and prevent HIV-1 replication in cell culture.² Recently, novel naphthyridines 2 and 3 were identified to be suitable replacements for the diketoacid pharmacophore.³ Furthermore, naphthyridine carboxamide 3 was found to be efficacious in rhesus macaques infected with the simian-human immunodeficiency virus (SHIV) 89.6P.⁴ In this communication, we describe the discov-

Keywords: HIV; Integrase; Strand transfer inhibitors; Dihydroxypyridopyrazine-1, 6-dione.

ery, SAR, computer modeling, pharmacokinetic profile, and synthesis of a novel series of dihydroxypyridopyrazine-1,6-diones HIV-1 integrase inhibitors.

Incorporation of the dihydroxycarbonyl pharmacophore (in red) into a pyridinone scaffold led to the dihydroxypyridinone⁵ moiety on the right-hand portion of the bicyclic system **4**. Addition of the benzyl amide functionality (in blue) and imposing restrictions on the rotation of the amide side chain via a cyclic constraint (in magenta) led to the novel dihydroxypyridinone-carboxylic derivatives depicted as **4**. Literature searches revealed no precedent for the preparation of such structure (Fig. 1).

Among the three different ring constrained compounds prepared, compounds 6 and 7 were found to be significantly more active against HIV integrase than compound 5 (Table 1). Molecular modeling (MMFF)⁷ of the bicyclic core suggests that the amide carbonyl group in the constrained seven-membered ring compound 5 is not coplanar relative to the remaining pharmacophore (Fig. 2), while in both 6-membered ring constrained analogs 6 and 7, the amide groups are coplanar with respect to the dihydroxypyridinone core.

^{*}Corresponding author. Tel.: +1 215 652 3020; e-mail: john_wai@merck.com

Figure 1. Conception of dihydroxypyridopyrazine-1,6-diones inhibitors.

Table 1. Effect of different constraints

Compound	X	Inhibition of strand transfer IC_{50}^{a} (μ M)
5	CH ₂ CH ₂ CH ₂	1.04 (±0.05)
6	CH_2CH_2	$0.10~(\pm 0.06)$
7	СН=СН	0.04 (±0.02)

 $[^]a$ Assays were performed with recombinant HIV-integrase (0.1 $\mu M)$ preassembled on immobilized oliogonucleotides. 6 Values are means of three experiments, standard deviation is given in parentheses.

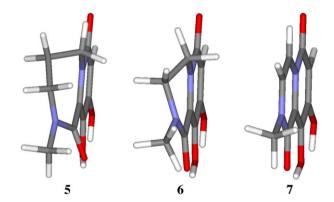


Figure 2. Molecular modeling of bicyclic core of inhibitors 5–7.

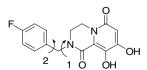


Figure 3. Torsional search on compound 6.

It was anticipated that the cyclic constraint would also bias the orientation of the benzyl side chain through steric interaction. Torsional search on compound 6 (varying the two bonds identified as 1 and 2 in Fig. 3) revealed four low energy regions. Two of the minima correspond to rotations about the 4-fluorophenyl ring

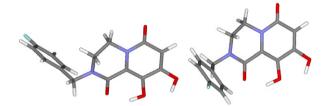


Figure 4. Two low energy conformers of compound 6.

Table 2. Effect of substitutions on benzyl group

Compound	R	Inhibition of strand transfer IC ₅₀ ^a (μM)	Antiviral activity in cell culture, CIC ₉₅ ^b (μM)
6	4-F	0.10 (±0.06)	0.31 (±0.08)
8	H	$0.23 (\pm 0.06)$	2.50 (±0.12)
9	2-C1	$0.37 (\pm 0.10)$	>1.00
10	3-C1	$0.04 (\pm 0.01)$	$0.50 (\pm 0.11)$
11	4-C1	$0.38 (\pm 0.07)$	1.00 (±0.18)
12	4-F, 3-Cl	$0.04 (\pm 0.02)$	$0.25~(\pm 0.03)$

^a Assays were performed with recombinant HIV-integrase (0.1 μM) preassembled on immobilized oliogonucleotides. ⁶ Values are means of three experiments, standard deviation is given in parentheses.

(bond 2). The two lower energy conformers differ only by rotation about bond 1 and are depicted in Figure. 4. This suggests that the bicyclic dihydroxypyrido-pyrazine-1,6-dione significantly limited conformation flexibility of the 4-fluorobenzyl side chain in compound 6.

Compound 6 inhibits 95% of the replication of HIV-1 in cell culture at $0.31 \,\mu\text{M}$ (Table 2). There is no shift in potency when the compound is assayed in the presence of 50% normal human serum.⁸ Removal of the 4-fluoro substitution in compound 6 leads to a significant drop in

^b Cell culture inhibitory concentrations (CIC₉₅) are defined as those which inhibited by > 95% the spread of HIV-1 infection in MT-4 human T-lymphoid cells maintained in RPMI 1640 medium containing 10% heat-inactivated fetal bovine serum. ⁸ Cytotoxicity is not observed in cell culture at concentrations up to 20 μM.

potency (cf. compound 8). A chloro substitution walk around the benzyl group shows that a 3-chloro substituent improves integrase inhibition relative to the 2- and 4-position isomers (compounds 10 vs 9 and 11). However, combining both the 4-fluoro and 3-chloro substituents only leads to a modest improvement in inhibitory potency in the cell based assay (compound 12). These compounds do not exhibit cytotoxicity in cell culture at concentrations up to $20~\mu M$.

In our attempt to attenuate the acidity of the 8-hydroxyl group to mimic diketoacid 1, electron-withdrawing substituents were incorporated adjacent to the 8-hydroxyl group on the bicyclic system. This led to a significant improvement in their intrinsic potency (Table 3, compounds 13, 14, 15 vs 6). However, this did not translate into improvements in antiviral activity. One possible explanation is that the equilibrium between neutral and ionized inhibitor has shifted to an extent which adversely affects the cell permeability of these analogs.

The bioavailability of compound 6 was 69% when dosed orally in rats at 10 mg/Kg as a solution in 1% aq meth-

Table 3. Effect of substitutions on pyridopyrazine-1,6-dione core

Compound	R′	Inhibition of strand transfer IC_{50}^{a} (μM)	Antiviral activity in cell culture, CIC ₉₅ ^b (μM)
6	Н	0.10 (±0.06)	0.31 (±0.08)
13	CN	$0.02 (\pm 0.06)$	>1.00
14	Br	$0.03 (\pm 0.01)$	$1.00 (\pm 0.16)$
15	I	$0.02 \ (\pm 0.02)$	>1.00

^a Assays were performed with recombinant HIV-integrase (0.1 μM) preassembled on immobilized oliogonucleotides. Values are means of three experiments, standard deviation is given in parentheses.

^b Cell culture inhibitory concentrations (CIC₉₅) are defined as those which inhibited by > 95% the spread of HIV-1 infection in MT-4 human T-lymphoid cells maintained in RPMI 1640 medium containing 10% heat-inactivated fetal bovine serum. ⁸ Cytotoxicity is not observed in cell culture at concentrations up to 20 μM.

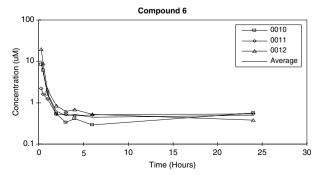


Chart 1. Oral pharmacokinetic profile of compound 6 in rat.

ylcellulose (Chart 1). Plasma concentrations of compound **6** were maintained between 0.64 and 0.50 μ M from the second to the twenty-fourth hour. This is well above the concentration (0.31 μ M) required for >95% inhibition of HIV-1 replication in cell culture.

There was a general concern about the biological liability of a dihydroxypyridinone core and its potential metabolites. ¹⁴C Labeled compound **6** was prepared⁹ to assess whether this series of compounds would irreversibly interact with liver microsomal proteins. Tables 4 and 5 depict the findings. There is only a low level of covalent interaction for radiolabeled **6** with liver microsomes derived from rat and human, well within the <50 pmol equiv/mg/60 min criteria (Table 4). ¹⁰ Furthermore, there is no difference in the results obtained with or without addition of NADPH. Results obtained in vivo in rat are also consistent with the observation made in vitro (Table 5).

The synthesis of compound 6 is depicted in Scheme 1. Reductive alkylation of dimethoxyethylamine with 4fluorobenzaldehyde 16 in the presence of sodium borohydride provided the corresponding benzylamine, which was treated with a mixture of N-acetylglycine, EDC, and HOBt in DMF to provide the bis amide 17. Acid catalyzed cyclization¹¹ of 17, followed by hydrogenation of the resultant product 18 in the presence of 5% Pd/C in ethanol, provided the acetyl piperazinone 19. Treatment of 19 in anhydrous DMF with LiHMDS, followed by addition of diethyl oxalate, led to three oxalation products, one major and two minor ones, as indicated by LC-MS analysis. Upon addition of more base, product 6 was generated from one of the minor oxalation products. The other two resisted cyclization and decomposed upon further exposure to base. The crude product mixture was purified by C-18 reverse phase HPLC eluting with a water-acetonitrile gradient.

Table 4. In vitro covalent binding of [14C] compound 6 with liver microsomes

Species	•	nd radioactivity /mg/60 min)
	-NADPH	+NADPH
Rat	24.7	24.2
Human	15.2	19.6

Table 5. In vivo covalent binding of [14C] Compound 6 in rats

Time (h)	•	Irreversibly bound radioactivity (pmol equiv/mg protein)	
	Plasma	Liver	
2	<5	<5	
6	<5	<5 <5 <5	
24	<5	<5	

Scheme 1. Synthesis of compound 6. Reagents: (a) i—H₂NCH₂CH(OMe)₂, NaBH₄, MeOH (82%); ii—*N*-Acetyl-Gly, EDC, HOBt, *i*-Pr₂NEt, DMF (95%); (b) MsOH, CH₂Cl₂ (62%); (c) H₂, 5% Pd/C, EtOH (95%); (d) i—LiHMDS, diethyl oxalate, DMF; ii—excess LiHMDS (15–20%).

NMR studies of the purified oxalation products led to the identification of the major component as 20, and one of the minor oxalation products as 22. Treatment of each one of them independently with LiHMDS in DMF did not lead to cyclization product 6. The third oxalation product readily cyclized to compound 6 upon workup. However, 20 was readily converted to the third oxalation product when exposed to a mixture of acetonitrile, water, and trifluoroacetic acid. NMR data of this material are consistent with those of structure 21. Treatment of 21 in DMF with LiHMDS led to the cyclization product 6.

In summary, a series of potent dihydroxypyrido-pyrazine-1,6-dione HIV-1 integrase inhibitors which inhibited replication of HIV-1 in cell culture has been

identified. The lead compound 6 inhibits replication of HIV-1 in cell culture at $CIC_{95} = 0.31 \,\mu\text{M}$, and exhibits no significant shift in potency when assayed with addition of 50% normal human serum. No cytotoxicity is observed in cell culture at concentrations up to 20 μM . When compound 6 was dosed orally in rat at 10 mg/kg as a solution in 1% methylcellulose, it displayed good pharmacokinetic profile and maintained excellent exposure through 24 h of the study. No covalent binding of compound 6 with microsomal proteins was observed in both vitro and in vivo models. Further exploration of this dihydroxypyridopyrazine-1,6-dione template and analogous bicyclic systems is in progress.

References and notes

- For a recent review on the structure and function of HIV-1 integrase, see: (a) Davies, D. R.; Chiu, T. K. Curr. Top. Med. Chem. 2004, 4, 965; (b) Pommier, Y.; Johnson, A. A.; Marchand, C. Nat. Rev. Drug Discov. 2005, 4, 236; For a recent review on HIV-1 integrase inhibitors, see: Gordon, C. P.; Griffith, R.; Keller, P. A. Med. Chem. 2007, 3, 199; Deng, J.; Dayam, R.; Al-Mawsawi, L. Q.; Neamati, N. Curr. Pharm. Des. 2007, 13, 129; Anthony, N. J. Curr. Top. Med. Chem. 2004, 4, 979.
- Wai, J. S.; Egbertson, M. S.; Payne, L. S.; Fisher, T. E.; Embrey, M. W.; Tran, L. O.; Melamed, J. Y.; Langford, H. M.; Guare, J. P., Jr.; Zhuang, L.; Grey, V. E.; Vacca, J. P.; Holloway, M. K.; Naylor-Olsen, A. M.; Hazuda, D. J.; Felock, P. J.; Wolfe, A. L.; Stillmock, K. A.; Schleif, W. A.; Gabryelski, L. J.; Young, S. D. J. Med. Chem. 2000, 43, 4923.
- 3. For monocyclic pyrimidinone integrase inhibitors, see (a) Zhuang, L.; Wai, J. S.; Embrey, M. W.; Fisher, T. E.; Egbertson, M. S.; Payne, L. S.; Guare, J. P., Jr.; Vacca, J. P.; Hazuda, D. J.; Felock, P. J.; Wolfe, A. L.; Stillmock, K. A.; Witmer, M. V.; Moyer, G.; Schleif, W. A.; Gabryelski, L. J.; Leonard, Y. M.; Lynch, J. J., Jr.; Michelson, S. R.; Young, S. D. J. Med. Chem. 2003, 46, 453; (b) Hazuda, D. J.; Anthony, N. J.; Gomez, R. P.; Jolly, S. M.; Wai, J. S.; Zhuang, L.; Fisher, T. E.; Embrey, M. W.; Guare, J. P., Jr.; Egbertson, M. S.; Vacca, J. P.; Huff, J. R.; Felock, P. J.; Witmer, M. V.; Stillmock, K. A.; Danovich, R.; Grobler, J.; Miller, M. D.; Espeseth, A. S.; Jin, L.; Chen, I.-W.; Lin, J.; Kassahun, K.; Ellis, J. D.; Wong, B. K.; Xu, W.; Pearson, P. G.; Schleif, W. A.; Cortese, R.; Emini, E.; Summa, V.; Holloway, M. K.; Young, S. D. Proc. Natl. Acad. Sci. U.S.A. 2004, 101, 11233.
- Hazuda, D. J.; Young, S. D.; Guare, J. P., Jr.; Anthony, N. J.; Gomez, R. P.; Wai, J. S.; Vacca, J. P.; Handt, L.; Motzel, S. L.; Klein, H. J.; Dornadula, G.; Danovich, R. M.; Witmer, M. V.; Wilson, K. A. A.; Tussey, L.; Schleif, W. A.; Gabryelski, L. S.; Lin, J.; Miller, M. D.; Casimiro, D. R.; Emini, E. A.; Shiver, J. W. Science 2004, 305, 528– 532
- (a) Summa, V.; Petrocchi, A.; Matassa, V.; Gardelli, C.; Muraglia, E.; Rowley, M.; Paz, O. G.; Laufer, R.; Monteagudo, E.; Pace, P. J. Med. Chem 2006, 49, 6646;
 (b) Petrocchi, A.; Koch, U.; Matassa, V. G.; Pacini, B.; Stillmock, K. A.; Summa, V. Biorg. Med. Chem. Lett. 2007, 17, 350.
- Hazuda, D. J.; Felock, P.; Hastings, J. C.; Pramanik, B.; Wolfe, A. J. Virol. 1997, 71, 7005, Assays were performed with recombinant HIV-1 integrase (0.1 μM) preassembled on immobilized oligonucleotides. Inhibitors were either

- added during assembly without washing or subsequent to assembly and washings. Inhibition was determined in relation to the integrase control reaction (without inhibitor) performed in quadruplicate and averaged. All samples were background subtracted.
- 7. Spartan '04 Windows ES 2.0.0. 2003.
- Vacca, J. P.; Dorsey, B. D.; Schleif, W. A.; Levin, R. B.; McDaniel, S. L.; Darke, P. L.; Zugay, J.; Quintero, J. C.; Blahy, O. M.; Roth, E.; Sairdana, V. V.; Schlabac, A. J.; Graham, P. I.; Condra, J. H.; Gotlib, L.; Holloway, M. K.; Lin, J.; Chen, I.-W.; Vastag, K.; Ostovic, D.; Anderson, P. S.; Emini, E. E.; Huff, J. R. Proc. Natl. Acad. Sci. U.S.A. 1994, 91, 4096–4100, 95% Cell culture inhibitory concentrations (CIC₉₅) are defined as those which inhibited by ≥95% the spread of HIV-1 infection in susceptible cell culture. MT-4 human T-lymphoid cells were main-
- tained in RPMI 1640 medium containing 10% heat inactivated fetal bovine serum. Cells were infected en masse at low multiplicity (0.01) using HIV-1 strain IIIb and were incubated for 24 h. At this time, cells were washed and distributed into 96-well microtiter dishes. Serial 2-fold dilutions of inhibitor were added to the wells and the cultures were maintained for three additional days. Virus spread was assessed by HIV-1 p24 core antigen ELISA. Control cultures in the absence of inhibitor were fully infected at 4 days.
- 9. ¹⁴C-labeled compound **6** was prepared as described in Scheme 1 using ¹⁴C-labeled diethyl oxalate.
- Evans, D. C.; Watt, A. P.; Nicoll-Griffith, D. A.; Baillie, T. A. Chem. Res. Toxicol. 2004, 17, 3–16.
- Kim, J. H.; Lee, Y. S.; Kim, S. S. Heterocycles 1998, 48, 2279–2286.